

**INHIBITORY EFFECT OF LINALOOL IN PREPARATIONS OF ISOLATED  
SMOOTH MUSCLE OF RAT TRACHEA WITH EPITHELIUM  
STIMULATED BY ELECTROMECHANICAL COUPLING**

*(Efeito inibitório do linalol em preparações isoladas de músculo liso traqueal de rato  
com epitélio estimulado por acoplamento eletromecânico)*

João Alison de Moraes Silveira<sup>1</sup>; Davi Sousa Rocha<sup>2</sup>; Glayciane Bezerra de Moraes<sup>3</sup>;  
Rodrigo José Bezerra de Siqueira<sup>1</sup>; José Henrique Leal-Cardoso<sup>2</sup>;  
Janaina Serra Azul Monteiro Evangelista<sup>3</sup>

<sup>1</sup>Faculdade de Medicina, Dptº de Fisiologia e Farmacologia - Universidade Federal do Ceará; <sup>2</sup>Instituto Superior de Ciências Biomédicas (UECE); <sup>3</sup>Faculdade de Veterinária, Universidade Estadual do Ceará (UECE), Av. Dr. Silas Munguba, 1700 - Campus do Itaperi, Fortaleza, Ceará.  
CEP: 60.714-502, Brazil.

**ABSTRACT**

Plants have been used as source of therapeutic elements employed to treat several disorders, such as hypertension and asthma. The discovery of pharmacological agents that act on the contractility of airways smooth muscle can be considered to help in treating diseases of the respiratory tract. In this study, we evaluated the effect of linalool, a terpenic constituent of various aromatic and medicinal plants as an antispasmodic agent in preparations of isolated rat trachea. Linalool fully reversed the electromechanical induced contraction in isolated preparations with (188.46±13.05 µM) and without (135.21±12.76 µM) epithelium. A lower inhibitory potency was observed in phamaco mechanical contractions induced in tracheal rings with preserved epithelium (261.34±38.22 µM) or not (593.45±11.32 µM). Linalool (100 µM and 1000 µM) was able to inhibit the curves of controlled influx for calcium and barium. In the present study, the inhibitory effect of linalool on pre-contracted rat tracheal rings was more effective in electromechanical conditions, suggesting a possible effect of this monoterpene on calcium influx through voltage-dependent channels.

**Keywords:** Monoterpene, smooth muscle, trachea.

**RESUMO**

As plantas têm sido utilizadas como fonte de elementos terapêuticos empregados para tratar vários distúrbios, como hipertensão e asma. A descoberta de agentes farmacológicos que atuam sobre a contratilidade do músculo liso das vias aéreas pode ser útil no tratamento de doenças do trato respiratório. Neste estudo, avaliamos o efeito do linalol, um constituinte terpênico de várias plantas aromáticas e medicinais como agente antiespasmódico em preparações de traqueia de rato isolada.

\*Endereço para correspondência:  
janainaserrazul@gmail.com

O linalol reverteu completamente a contração eletromecânica induzida em preparações isoladas com (188,46±13,05 µM) e sem epitélio (135,21±12,76 µM). Observou-se uma menor potência inibitória nas contrações fármaco mecânicas induzidas em anéis traqueais com epitélio preservado (261,34±38,22 µM) ou não (593,45±11,32 µM). O linalol (100 µM e 1000 µM) foi capaz de inibir as curvas de influxo controlado para cálcio e bário. No presente estudo, o efeito inibitório do linalol em anéis traqueais de ratos pré-contraídos foi mais eficaz em condições eletromecânicas, sugerindo um possível efeito deste monoterpene sobre o influxo de cálcio através de canais dependentes da tensão.

**Palavras-chave:** Monoterpene, músculo liso, traqueia.

## INTRODUCTION

The essential oils of Brazilian plant biodiversity, such as *Aniba rosaeodora* (Rosewood) have the terpenic constituent linalool (ALMEIDA *et al.*, 2013; BAKKALI *et al.*, 2008). Linalool (LNL), or 3,7-dimethyl-1,6-octadien-3-ol is an acyclic tertiary monoterpene alcohol having two active chemical isomers. Biological effects of linalool-rich rosewood oil include sedative (DE ALMEIDA *et al.*, 2009), anticancer (SOEUR *et al.*, 2011) and cardiovascular actions (DE SIQUEIRA *et al.*, 2014). These cardiovascular effects are related to the induction in vivo of a vago-vagal reflex in addition to vasodilatory properties in isolated smooth muscle preparations of rat aorta (DE SIQUEIRA *et al.*, 2014).

The constituent linalool is able to relax the smooth muscle of isolated vascular preparations from rats (ANJOS *et al.*, 2013). This vaso relaxant effect was also observed in mouse aortic rings and

involves the participation of soluble guanylyl cyclase and K<sup>+</sup> channels (KANG and SEOL, 2015). Therefore, this study aimed to investigate the effects of linalool on smooth muscle preparations of rat trachea stimulated by electromechanical and pharmacomechanical coupling.

## MATERIAL AND METHODS

### Ethical Aspects

This study was conducted in compliance with the international rules established by the Guide for the Care and Use of Laboratory Animals and was submitted and approved by the Ethics Committee for the Use of Animals of State University of Ceará with the protocol number 10244898-1.

### Animals

In this study were used male Wistar rats (250±50g) from the local colonies maintained in the Federal

University of Ceará, Fortaleza, Brazil. These animals were maintained in polypropylene boxes, at a temperature of  $24 \pm 2$  °C, fed with ration and water *ad libitum*, and at a light-dark cycle of 12 hours.

### **Isolated Preparations of Trachea Smooth Muscle**

Animals were euthanized by pentobarbital anesthesia (50 mg/kg, i.p.) followed by fast aorta exsanguination. The trachea were dissected, isolated in Petri dishes with nutrient solution and divided in sections of 5 mm length allocated in steel triangular devices (0.3 mm diameter) coupled with isometric force transducers (ML870B60/C-V, ADInstruments, Bella Vista, Australia) and a data acquiring system (PowerLab 8/30, ADInstruments, Bella Vista, Australia). The tracheal rings were placed in 5 mL organ baths of Tyrode at 36.5 °C and continuously bubbled with a 95% O<sub>2</sub> and 5% CO<sub>2</sub> air mixture generated by a perfusion system. The epithelium mechanical denudation was performed with a slight friction in the lumen of the tracheal ring and confirmed after histological analysis.

### **Experimental Protocols**

**Series 1:** Preparations with or without epithelium were stimulated by the

electromechanical (added KCl 80 mM) or pharmacomechanical (added carbachol 1 μM) coupling. Once identified the maximal contraction, cumulative concentrations of LNL (1 to 3000 μM) were added to the preparation. After the LNL application, Tyrode washings were performed in every 15 min. during the recuperation period of 60 min or longer, if necessary. Only then, a new contraction was stimulated with KCl 80 mM, in order to assess the tissue response after the protocol.

**Series 2:** In another experimental series designed to reinforce the mechanism of action of LNL related to voltage-operated calcium channels (VOCC), tracheal rings with epithelium were maintained in Ca<sup>2+</sup>-free medium in the presence of KCl (80 mM) and EGTA (1 mM). After 20 min Ca<sup>2+</sup> (0.1 to 10 mM) was cumulatively added to the organ bath in the absence or presence of LNL (10 μM, 100 μM and 1000 μM). In another group of preparations, Ca<sup>2+</sup> was replaced by the cumulative addition of Ba<sup>2+</sup> (0.1 to 10 mM).

### **Solutions and Drugs**

All the salts and substances necessary for this study, and the LNL in its racemic form as well, were acquired from Sigma Chemical Corporation (St. Louis, USA) and Reagen (Rio de Janeiro,

BRAZIL). Tissues were maintained in modified Tyrode preparation composed of (in mM) NaCl 136.0; KCl 5.0; MgCl<sub>2</sub> 0.98; NaH<sub>2</sub>PO<sub>4</sub> 0.36; NaHCO<sub>3</sub> 11.9; CaCl<sub>2</sub> 2.0; glucose 11.0, with the due modifications necessary for the concentration-effect ratio experiments of calcium (Ca<sup>2+</sup>) and barium (Ba<sup>2+</sup>). The pH of the solution was stabilized in 7.4 (37 °C; continuous bubbling with 5% CO<sub>2</sub> in 95% O<sub>2</sub>) before the experiments. The LNL was dispersed and homogenized in nutrient solution applied in the protocols added Tween 80 in a 0.5% proportion. The other substances were diluted or dispersed in distilled water.

### Statistical Analysis

The results were expressed as mean ± SEM and n indicates the number of experiments. For each protocol, the IC<sub>50</sub> (i.e., the concentration of LNL at which 50% of a contractile response was inhibited) values were calculated by logarithm interpolation. Paired or unpaired Student's t-tests and One-way ANOVA followed by Bonferroni *post-hoc* test were used for comparing, respectively, two groups and more than two groups. Statistical significance was accepted at p<0.05. The data were analyzed using Sigma Plot 10 (Systat Software Inc., Chicago, USA).

## RESULTS AND DISCUSSION

The LNL, but not the vehicle, was able to reverse the contraction evoked by KCl-induced electromechanical coupling (Fig. 1). According to the results, both preparations with the preserved epithelium (Fig. 1-A) as well as in the ones where this tissue was removed (Fig. 1-B), the LNL was capable of reversing the contraction. In both preparations, the relaxing occurred significantly from the concentration of 100 µM, achieving complete reversion at 600 µM of LNL. There was no statistically significant difference in IC<sub>50</sub> values when the reversals of the electromechanical coupling of epithelial (188.46±13.05 µM) and non-epithelial (135.21±12.76 µM) preparations were compared.

The contraction induced by the pharmacomechanical coupling was reversed by LNL, but not by its vehicle (Fig. 2). For reversal of carbachol precontraction, IC<sub>50</sub> was significantly lower (261.34±38.22 µM) in preparations with epithelium compared to tracheal preparations epithelium-denuded (593.45±11.32 µM). With the pharmacomechanical induction, the results also presented contraction reversal in the preparations with (Fig. 2-A) or without the epithelial tissue (Fig. 2-B).

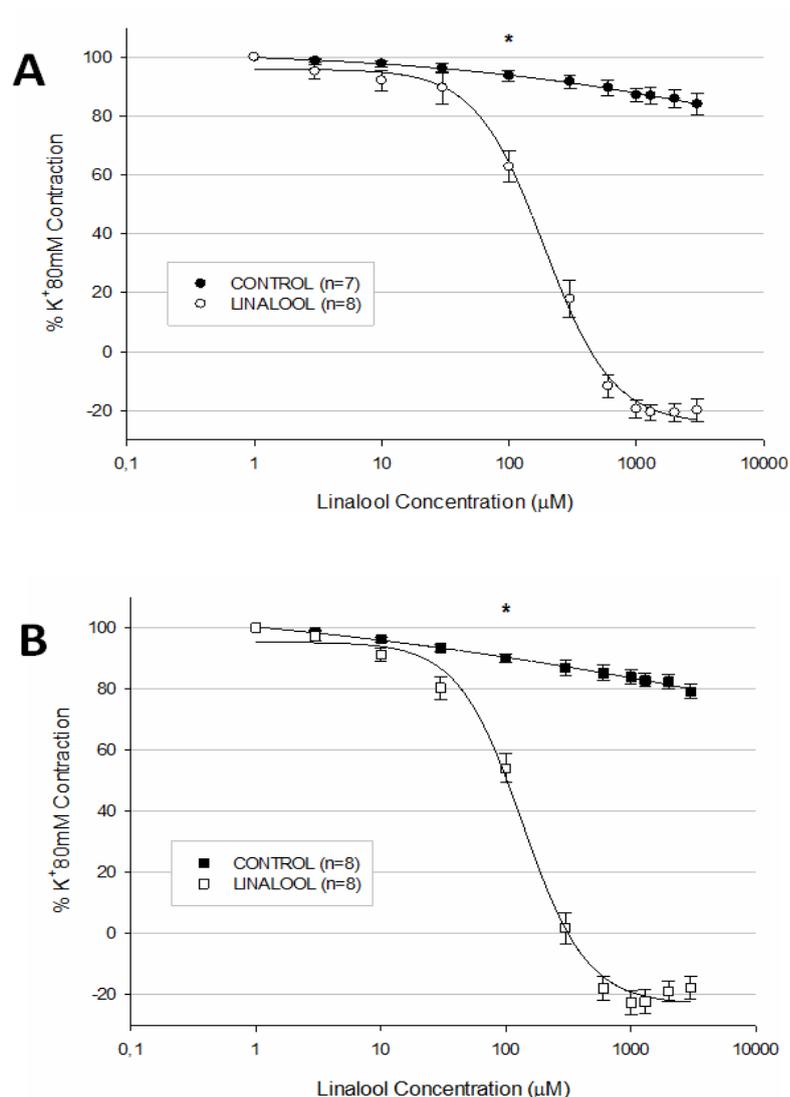


Figure 1: Antispasmodic effect of growing and cumulative concentrations (1 to 3000μM) of LNL on the contraction induced by K<sup>+</sup>80mM of tracheal rings isolated from rat.

The data are presented as means±SEM (p<0.05). (A) Preparation with the preserved epithelium and; (B) Preparation without epithelium.

Statistical significance was observed between the preparations. In the experiment with the epithelium, the concentration of 30 μM firstly demonstrated significant relaxing, but the reversal, unlike the other experiments, was not complete (2.43±2.73%). In the experiments without the epithelium, the relaxing was not significant until the

concentration of 300 μM was achieved, reaching full reversal at the concentration of 2000 μM.

In the concentration-response curves of Ca<sup>2+</sup> (Fig. 3-A), only the 10 μM LNL concentration did not have a significant difference when compared to control, even though the curve was dislocated to the right. The concentrations

of 100  $\mu\text{M}$  and 1000  $\mu\text{M}$  were significantly different in reducing the effect of the cumulative concentrations of

$\text{Ca}^{2+}$ , always in comparison to control group.

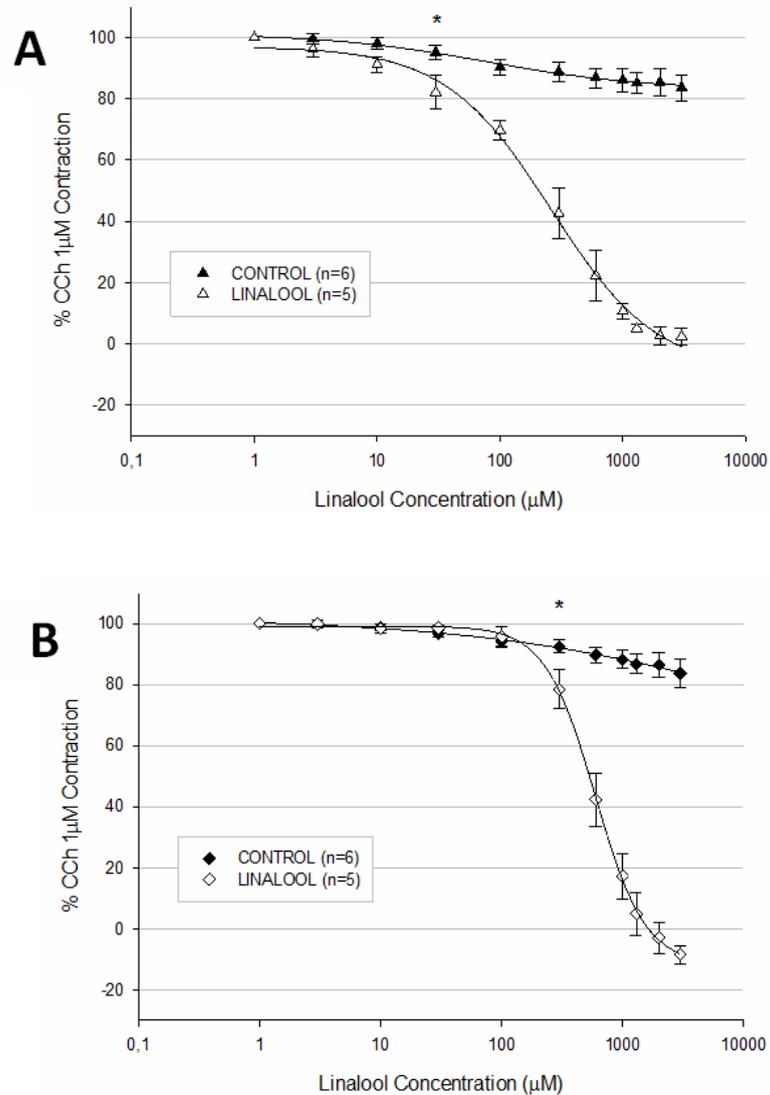


Figure 2: Antispasmodic effect of growing and cumulative concentrations (1 to 3000 $\mu\text{M}$ ) of LNL on the contraction induced by CCh 1 $\mu\text{M}$  of tracheal rings isolated from rat. The data is presented as means $\pm$ SEM.

(\*) Concentration that presented significant difference ( $p < 0.05$ ) between control and experimental group. (A) Preparation with the preserved epithelium and (B) preparation without epithelium.

Occurrences on the concentration-response curves for  $\text{Ca}^{2+}$  were similar to those observed in the protocol for  $\text{Ba}^{2+}$  (Fig. 3-B). The

difference observed was in the greater dislocation to the right in the concentration of 10  $\mu\text{M}$  LNL used, with the statistical difference found until the 3

mM  $Ba^{2+}$  concentration, reaching similar results as the control group in the final concentrations. Other LNL concentrations (100  $\mu$ M and 1000 $\mu$ M) were able to inhibit the contraction after adding more  $Ba^{2+}$ . Even without significant difference

when the effects of the same concentrations between the  $Ca^{2+}$  and  $Ba^{2+}$  were compared, it appeared that the blocking in the latter ion curve was greater.

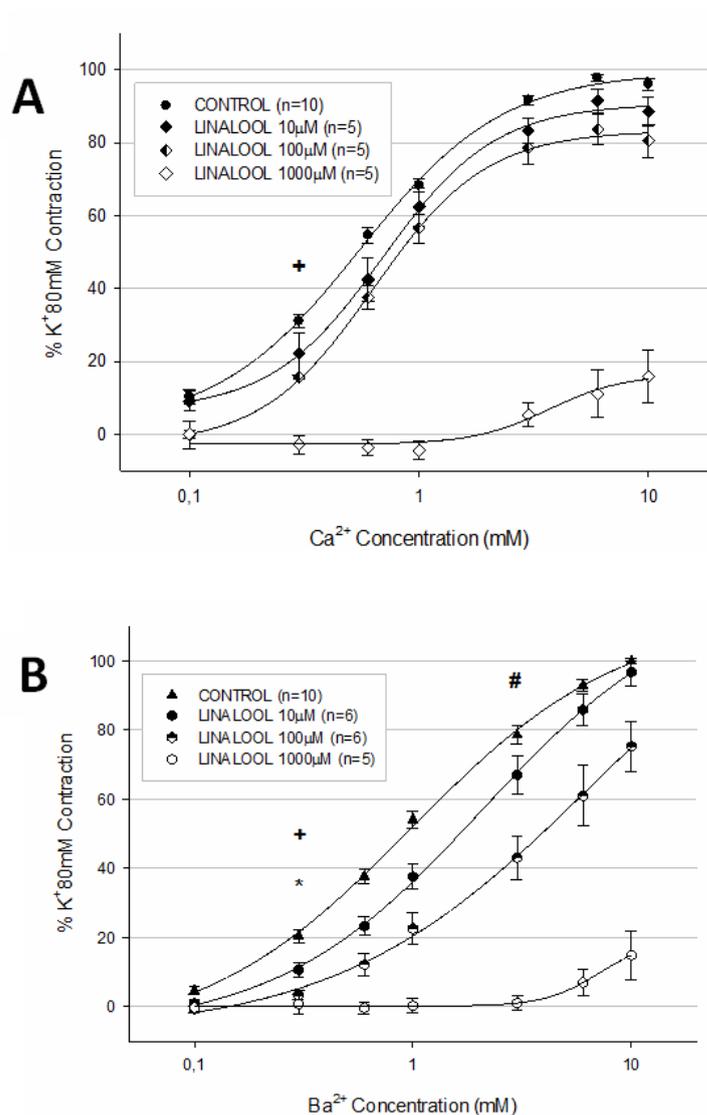


Figure 3: Effect of three concentrations (10, 100 and 1000 $\mu$ M) of LNL on the growing and cumulative concentrations (0.1 to 10mM) of  $Ca^{2+}$ (A) and  $Ba^{2+}$ (B) on the contraction induced by  $K^{+}80$  of the tracheal rings isolated from rat.

The data is presented as means $\pm$ SEM. (\*-#) Interval of concentrations that presented significant difference ( $p < 0.05$ ) between the control and experimental group of 10 $\mu$ M LNL. (+) Concentration that presented significant difference ( $p < 0.05$ ) between control and experimental group of 100 $\mu$ M and 1000 $\mu$ M LNL.

We could infer that the LNL presented greater relaxant effect in the inductions by electromechanical induction than in those with the pharmacomechanical contraction since the  $IC_{50}$  obtained from the preparations induced by  $K^+$  80mM were lower than the ones achieved with the CCh stimulation. In addition, statistical differences were observed in comparing the  $IC_{50}$  found in both types of induction in the presence of epithelium which was also observed when that tissue was removed.

The differences found between the pharmacological potencies in the results of electromechanical and pharmacomechanical protocols, may have occurred because ligand stimulation activates intrinsic mechanisms, such as  $IP_3$ , cADPR and the RHOK route, besides the opening of calcium membrane channels producing a more sustainable contraction (GOSENS *et al.*, 2006; MCFADZEAN and GIBSON, 2002). If this premise endure, and according to the effect on the electromechanical coupling discovered, we might conclude that the LNL did not possess a blocking activity on the muscarinic receptors.

When the most probable mechanisms involving the tracheal smooth muscles were stipulated, the true participation of the epithelium in the muscle relaxing effect of LNL could not

be concluded. In the protocols of contraction induced by  $K^+$  80, the preparations in which the epithelium was removed achieved more potent values, than those in which this tissue was preserved, while in the CCh stimulations, the opposite effect was observed.

Based on the results obtained and that the epithelium releases factors that may act in specific receptors, such as endothelin and gases such as NO (GOLDIE *et al.*, 1990; JANSSEN and KILIAN, 2006), this tissue may even potentiate the antispasmodic activity of the LNL in the organism. How the compound acts on the epithelium to achieve the relaxing effects could not be asserted. Actually, since the significant reversal was observed in all four previous situations, we might suggest that the LNL acts directly on the smooth muscle, activating processes that initiate the muscle relaxing or blocking the ionic channels.

This may have occurred due to a blocking in the  $K^+$  channels, often observed in elevated concentrations of  $Ba^{2+}$  ion. However, the elevated inhibition in both curves – when stimulated by  $K^+$  80 – suggested that LNL had an effect on ion flux VOCC type  $Ca^{2+}$  channels, activated by electromechanical coupling and the only by which the  $Ba^{2+}$  ions permeate

(JANSSEN *et al.*, 2004; MURRAY and KOTLIKOFF, 1991).

In conclusion, LNL is capable of reversing the pre-contraction induced by different agents in isolated tracheal preparations. The difference between the two IC<sub>50</sub> was significant and an elevated dislocation to the right of the preparation without epithelium occurred, indicating that, for this coupling form, the participation of the epithelium-derived factor is important for the relaxing promotion. This inhibitory effect of LNL was more potent in preparations on electromechanical challenge. These findings suggest that linalool interfere with membrane calcium influx through voltage-dependent channels.

#### **CONFLICT OF INTEREST**

The authors of this article declare that there is no potential conflicts of interest including employment, consultancies, stock ownership, honoraria, paid expert testimony and patent applications/registrations related to the current manuscript. This manuscript is submitted on behalf of all authors.

#### **ACKNOWLEDGMENTS**

We would like to thank CNPq, CAPES, FINEP and FUNCAP by the financial support of this research. And to the professor Dr. Pedro Jorge Caldas Magalhães, for the great scientific collaboration, for lending the facilities in which the experiments were performed and for the help with all the teaching, advising and discussing of the obtained results.

#### **REFERENCES**

- ALMEIDA, M.R.; FIDELIS, C.H.; BARATA, L.E., POPPI, R.J. 2013. Classification of Amazonian rosewood essential oil by Raman spectroscopy and PLS-DA with reliability estimation. *Talanta*, v.15, n.117, p.305-311, 2013.
- ANJOS, P.J.; LIMA, A.O.; CUNHA, P.S.; DE SOUSA, D.P.; ONOFRE, A.S.; RIBEIRO, T.P.; MEDEIROS, I.A.; ANTONIOLLI, A.R.; QUINTANS-JÚNIOR, L.J.; SANTOSA, M.R. Cardiovascular effects induced by linalool in normotensive and hypertensive rats. *Zeitschrift für Naturforschung C*, v.68, p.181-190, 2013.
- BAKKALI, F.; AVERBECK, S.; AVERBECK, D.; IDAOMAR, M. Biological effects of essential oils – A

- review. Food and Chemical Toxicology, v.46, n.2, p. 446-475, 2008.
- COELHO-DE-SOUZA, A.N.; BARATA, E.L.; MAGALHÃES, P.J.C.; LIMA, C.C.; LEAL-CARDOSO, J.H. Effects of the essential oil of *Croton zehntneri*, and its constituent estragole on intestinal smooth muscle. Phytotherapy Research, v.11, p.299-304, 1997.
- DE ALMEIDA, R.N.; ARAÚJO, D.A.; GONÇALVES, J.C.; MONTENEGRO, F.C.; DE SOUSA, D.P.; LEITE, J.R.; MATTEI, R.; BENEDITO, M.A.; DE CARVALHO, J.G.; CRUZ, J.S.; MAIA, J.G. Rosewood oil induces sedation and inhibits compound action potential in rodents. Journal of Ethnopharmacology, v.30, n.124, p.440-443, 2009.
- DE SIQUEIRA, R.J.; RODRIGUES, K.M.; DA SILVA, M.T.; CORREIA JUNIOR, C.A.; DUARTE, G.P.; MAGALHÃES, P.J.; DOS SANTOS, A.A.; MAIA, J.G.; DA CUNHA, P.J.; LAHLOU, S. 2014. Linalool-rich rosewood oil induces vago-vagal bradycardic and depressor reflex in rats. Phytotherapy Research, v.28, n.1, p.42-48.
- ESTEVES, I.; SOUZA, I.R.; RODRIGUES, M.; CARDOSO, L.G.V.; SANTOS, L.S.; ABOIN-SERTIE, J.A.; PERAZZO, F.F.; LIMA, L.M.; SCHNEEDORF, J.M.; BASTOS, J.K.; CARVALHO, J.C.T. Gastric antiulcer and anti-inflammatory activities of the essential oil from *Casearia sylvestris* Sw. Journal of Ethnopharmacology, v.101, p.191-196, 2005.
- EVANGELISTA, G.L.; COELHO-DE-SOUZA, A.N.; SANTOS, C.F.; LEAL-CARDOSO J.H.; LOPES, E.A.; SANTOS, M.V.; LAHLOU, S.; MAGALHÃES, P.J.C. 2007. Essential oil of *Pterodon olygalaeflorus* inhibits electromechanical coupling on rat isolated trachea. Journal of Ethnopharmacology, v.109, n.3, p.515-522.
- GOLDIE, R.G.; FERNANDES, L.B.; FARMER, S.G.; HAY, D.W.P. Airway epithelium-derived inhibitory factor. Trends Pharmacological Sciences, v.11, p.67-70, 1990.
- GOSENS, R.; ZAAGSMA, J.; MEURS, H.; HALAYKO, A.J. Muscarinic receptor signaling in the pathophysiology of asthma and COPD. Respiratory Research, v.7, n.73, p.1-15, 2006.
- JANSSEN, L.J.; KILIAN, K. Airway smooth muscle as a target of asthma therapy: history and new directions. Respiratory Research, v.7, n.123, p.1-12, 2006.

JANSSEN, L.J.; TAZZEO, T.; ZUO, J.; PERTENS, E.; KESHAVJEE, S. KCl evokes contraction of airway smooth muscle via activation of RhoA and Rho-kinase. *American Journal of Physiology Lung Cell Molecular Physiology*, v.287, p.852-858, 2004.

KANG, P.; SEOL, G.H. Linalool elicits vasorelaxation of mouse aortae through activation of guanylyl cyclase and K(+) channels. *Journal of Pharmacy and Pharmacology*, v.67, n.5, p.714-719, 2015.

LIMA-ACCIOLY, P.M.; LAVOR-PORTO, P.R.; CAVALCANTE, F.S.; MAGALHÃES, P.J.C.; LAHLOU, S.; MORAIS, S.M.; LEAL-CARDOSO, J.H.. Essential oil of *Croton nepetaefolius* and its main constituent, 1,8-cineole, block excitability of rat sciatic nerve *in vitro*. *Clinical Experimental Pharmacology and Physiology*, v.33, n.12, p.1158-1163, 2006.

MCFADZEAN, I.; GIBSON, A. The developing relationship between receptor-operated and store-operated calcium channels in smooth muscle. *British Journal of Pharmacology*, v.135, n.1, p.1-13, 2002.

MURRAY, R.K.; KOTLIKOFF, M.I. Receptor-activated calcium influx in

human airway smooth cells. *Journal of Physiology*, v.435, p.123-144, 1991.

SOEUR, J.; MARROT, L.; PEREZ, P.; IRAQUI, I.; KIENDA, G.; DARDALHON, M.; MEUNIER, J.R.; AVERBECK, D.; HUANG, M.E.

Selective cytotoxicity of *Aniba rosaeodora* essential oil towards epidermoid cancer cells through induction of apoptosis. *Mutation Research*, v.10, n.718, p.24-32, 2011.